

S0040-4039(96)00453-4

Phœniceroside, the First Natural Bis-Furanone Propane Derivative from *Juniperus phœnicea* L.

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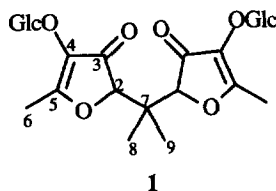
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Abstract: Phœniceroside **1** or 2,2-bis-(5-methyl-4-(*O*-β-D-glucopyranosyl)-3(2H)-furanone)-propane have been isolated from *Juniperus phœnicea* L. (Cupressaceae). The structural elucidation of this new natural product was achieved by UV, MS, ¹H, ¹³C NMR spectroscopy. The structure was confirmed by chemical way. Copyright © 1996 Elsevier Science Ltd

Recently, a German team had isolated from a plant and identified psydrin, a furanone glucoside derivative¹. A few weeks later, we characterized the same compound from *Juniperus phœnicea* along with phœnicein, another new natural furanone derivative². These compounds are members of the furaneol family described to have mutagenic properties^{3,4}. In this paper, we report the isolation of a new dimer related to these molecules from the same *Juniperus* and its characterization by spectroscopy and chemical correlation.



The M_r 592 and the molecular formula $C_{25}H_{36}O_{16}$ for **1** were deduced from its MS spectrum. Full assignments of ¹H and ¹³C peaks, the connectivities between protons and carbons, as well as linkages remaining between glucosyl and aglycone units were established by 2D ¹H-¹H homonuclear correlation spectroscopy, *J* modulated ¹³C, HMQC and HMBC experiments (Table 1). The positive and negative FAB mass spectra⁵ showed respectively quasi-molecular ions at m/z 593 [M+H]⁺ and 591 [M-H]⁻. This was confirmed by the DCI and electrospray mass spectra⁵. The glucosyl moiety in the natural product **1** was revealed evidently in the ¹H NMR spectrum by the anomeric proton signal at δ 4.80 ($J = 8\text{Hz}$)⁶, by the pattern of the carbon signals⁷ and by the loss of 162 atomic mass units noted in the FAB mass spectra⁵. The glucosyl unit was also identified after acid hydrolysis of compound **1**. The linkage between the sugar residue and the aglycone part was given by the HMBC ³*J* correlation observed between the anomeric osidic proton and the ethylenic carbon at δ 136.3 (C-4) (Table 1).

	Phœniceroside			Psydrin		Phœnicein	
	¹ H	¹³ C	HMBC	¹ H	¹³ C	¹ H	¹³ C
2	4.85 <i>s</i>	86.5*	16.9; 43.0; 136.3; 184.2; 198.1	4.30 <i>s</i> (2H)	74.8		144.4
3		198.1			198.0		182.2
4		136.3			135.7		137.3
5		184.2			186.0		171.4
6	2.36 <i>s</i>	13.9	136.3; 184.2; 198.1	2.31 <i>s</i> (3H)	14.2	2.34 <i>s</i> (3H)	12.7
7		43.0*					137.0
8/9	1.10* <i>s</i>	16.9* 17.1*	16.9; 17.1; 43.0; 86.5			2.30 <i>s</i> (3H) 2.06 <i>s</i> (3H)	17.0/19.3
1'	4.80* <i>d</i> (8)	104.5*	78.4; 136.3	4.75 <i>d</i> (7.8)	104.5	4.74 <i>d</i> (7.9)	105.3
2'	3.27 <i>m</i>	74.8	77.8; 104.5	<i>ca</i> 3.32 <i>m</i>	74.8	<i>ca</i> 3.28 <i>m</i>	74.8
3'	3.39 <i>m</i>	77.8	71.3; 74.8; 78.4	<i>ca</i> 3.32 <i>m</i>	77.8	3.41 <i>t</i> (8.8)	76.8
4'	3.35 <i>m</i>	71.3	62.5; 77.8; 78.4	<i>ca</i> 3.32 <i>m</i>	71.3	3.35 <i>t</i> (9.5)	71.1
5'	3.25 <i>m</i>	78.4	62.5; 71.3; 77.8	<i>ca</i> 3.32 <i>m</i>	78.4	<i>ca</i> 3.30 <i>m</i>	78.5
6'	3.72 <i>m</i> 3.86 <i>m</i>	62.5	71.3; 78.4	3.67 <i>dd</i> (12; 5.3) 3.83 <i>dd</i> (12; 2.1)	62.6	3.69 <i>dd</i> (11.9; 5.3) 3.83 <i>dd</i> (11.9; 2.3)	62.6

Table 1 : ¹H and ¹³C NMR data of psydrin, phœnicein² and phœniceroside **1** (CD₃OD, 500 MHz)
(* multiple signals).

Both the ¹H and ¹³C NMR spectra of **1** showed some great similarities with those of psydrin and phœnicein we had already described (Table 1)². The most important difference was the appearance of a quaternary aliphatic carbon signal at δ 43.0 (C-7) instead of a quaternary ethylenic one in the phœnicein. The deshielding of H-2 (δ 4.85) and C-2 (δ 86.5) relative to psydrin was in agreement with the higher substitution of C-2. The good matching of the chemical shifts of C-3, C-4, C-5 and C-6 with the data for psydrin indicated the identity of structure for this part of the molecule. The molecular weight obtained from all MS data⁵ indicated that compound **1** was constituted by two furanone derivative units and the simplicity of the NMR spectra showed a relative symmetry in the structure. Final structural elucidation was given by the HMBC experiment. Actually, a ²J cross peak observed between H-2 and C-7 indicated the link between the quaternary carbon and the C-2. Moreover, the ³J cross peaks between H-2 and C-8/C-9, between H-8/H-9 and C-7 demonstrated the propane chain. Finally, a ³J correlation observed between the H-2 of a furanone unit and the C-2 of the other one gave proof of the pseudo-dimerization. Moreover, the chromatographic behaviour of disaccharide type comparatively to those of psydrin and phœnicein² was in agreement with a diglucoside structure. However, the multiplicity of ¹H and ¹³C signals for the positions 2, 8, 9, 7 and 1' indicated a mixture of isomers.

Finally, the acid hydrolysis of **1** (Figure 1) led to two purified derivatives **1a** and **1b** identified mass spectrometry and NMR spectroscopy^{8,9}. Actually, in DCI+MS showed in both cases quasi-molecular ions at m/z 286 $[M+NH_4]^+$, 269 $[M+H]^+$ and 268 $[M+NH_4-H_2O]^+$ indicated the molecular weight of hydrolyzed compounds at MM 268 in agreement with the structural hypothesis for **1**. Furthermore, the NMR spectra of these compounds showed the expected signals. The difference between the two hydrolyzed compounds was the multiplicity of C-8/H-8 and C-9/H-9. Compound **1a** showed methyls 8 and 9 as singlet at δ 17.1 corresponding to the relative *meso* form. Compound **1b** showed for the same methyls two signals at δ 16.5 and δ 17.8 corresponding to the *threo* form.

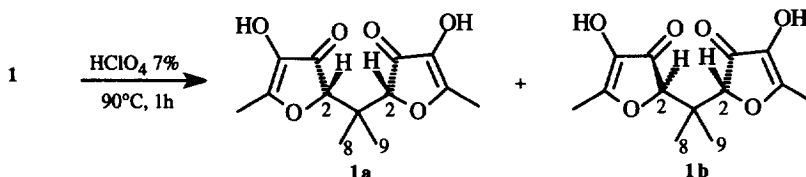


Figure 1 : Acid hydrolysis of phœniceroside **1**.

The structure of **1** was confirmed by the hemisynthetic compound obtained from the condensation of stoichiometric amounts of phœnicein and psydrin (Figure 2). The synthetic product had completely identical NMR data (¹H and ¹³C) and chromatographic behaviour (TLC and HPLC) as the natural one.

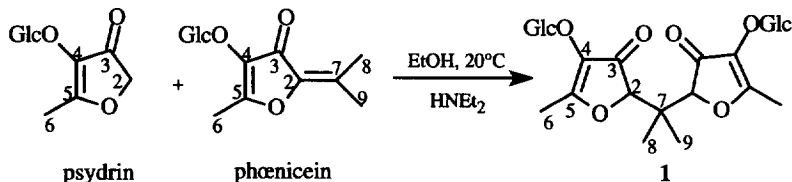


Figure 2 : Condensation of psydrin and phœnicein leading to compound **1**.

These experiments showed us that natural compound **1** was a mixture of diastereomers. The relative lability of the H-2 seemed to be at the origin of the isomerization. During the last 30 years, studies on volatile flavour compounds from a large spectra of fruit-producing plants led to the discovery of the very important family of the Furaneol™ and related compounds. The 2,5-dimethyl-4-hydroxy-3(2H)-furanone (furaneol) was first isolated from pineapple¹⁰. Some authors have reported the 4-methoxy-furaneol (mesifurane)¹¹, and recently the furaneol glucoside occurring as a mixture of diastereomers¹². We previously described in *Juniperus phœnicea* two other glucoside derivatives of this family, psydrin and phœnicein². The new phœniceroside seems to result of the condensation of the both compounds, but the question is asked to know if this mix is due to the plant or to our working methods using aqueous solutions during the purification. However, the very low efficiency (7%) of the hemisynthesis in connection with the relative amounts obtained for the three furanone derivatives during the purification indicated a biogenetic pathway for **1**. Consequently, it would be interesting to study the mutagenic potentialities of this new compound comparatively to those described for the furaneol^{3,4} even if antiproliferative tests on human tumoral and leucemic cell lines we have made on psydrin and phœnicein have not given significative results.

References and Notes

- Nahrstedt, A., Rockenbach, J. and Wray, V. *Phytochemistry*, **1995**, *39*, 375-378.
- Comte, G., Allais, D. P., Chulia, A. J., Vercauteren, J. and Delage, C. *Phytochemistry*, (MS 6532, in press).
- Tian, Q., Li, G., Shen, J. and Wang, Y. *Weisheng Dulixue Zazhi*, **1992**, *6*, 26-28.
- Xing, B., Liu, K., Li, Y., Yao, A., Li, Y., Zhi, X., Zhang, X. and Zheng, A. *Zhonghua Yufagyixue Zazhi*, **1988**, *22*, 85-87.
- phoenicoside 1. $[\alpha]_D^{20} = -80^\circ$ ($c = 0,15$; MeOH). UV $\lambda_{\max}^{\text{MeOH}}$ nm (ϵ): 279 (36700). Chromatographic mobilities were recorded in four systems : system 1 (Si gel F-254, EtOAc:HCOOH:HOAc:H₂O = 20:1:1:2) R_f 0.26, system 2 [cellulose F-254, *n*-BuOH:HOAc:H₂O = 4:1:5 (upper phase)] R_f 0.14, system 3 (cellulose F-254, HOAc:H₂O = 3:17) R_f 0.87, system 4 [LichroCART 250-4 Lichrospher 100 DIOL 5 μm , *n*-Hexane:MeOH:*i*-PrOH = 63:30:7, 1 ml.min⁻¹] R_t 12 min. ¹H and ¹³C NMR (see Table 1). DCI⁺MS (NH₃ + isobutane) : m/z 610 [M+NH₄]⁺, 593 [M+H]⁺, 317 [M-psydrin+H]⁺, 155 [M-psydrin-Glc+H]⁺, 115 [M-hydrophoenicein-Glc+H]⁺. Electrospray MS : m/z 615 [M+Na]⁺. EIMS : m/z 276 [M-hydrophoenicein]⁻, 154 [M-psydrin-Glc]⁻, 114 [M-hydrophoenicein-Glc]⁻. FAB⁺MS (glycerol + KCl) : m/z 632 [M+K]⁺, 593 [M+H]⁺, 431 [M-Glc+H]⁺, 317 [M-psydrin+H]⁺, 277 [M-hydrophoenicein+H]⁺, 155 [M-psydrin-Glc+H]⁺. FAB⁻MS (glycerol + KCl) : m/z 627 [M+Cl]⁻, 591 [M-H]⁻, 429 [M-Glc-H]⁻, 315 [M-psydrin-H]⁻, 275 [M-hydrophoenicein-H]⁻, 113 [M-hydrophoenicein-Glc-H]⁻.
- Perlin, A. S. and Casu, B. *Tetrahedron Letters*, **1969**, *34*, 2921-2924.
- Breitmayer, E. and Voelter, W. *Carbon-13 NMR Spectroscopy*. **1989**, VCH, Weinheim.
- Compound 1a. UV $\lambda_{\max}^{\text{MeOH}}$ nm : 289 . Chromatographic mobilities were recorded in two systems : system 5 (Si gel F-254, CHCl₃:MeOH = 9:1) R_f 0.45, system 6 [LichroCART 250-4 Lichrospher 100 DIOL 5 μm , *n*-Hexane:MeOH:*i*-PrOH = 18:5:2, 1 ml.min⁻¹] R_t 8.5 min. ¹H NMR (500 MHz in MeOH) : δ 1.05 (H-8/9), 2.19 (H-6), 4.63 (H-2). ¹³C NMR (125 MHz in MeOH) : 12.9 (C-6), 17.1 (C-8/9), 43.0 (C-7), 85.7 (C-2), 136.7 (C-4), 174.7 (C-5), 198.3 (C-3). DCI⁺MS (NH₃ + isobutane) (rel. int.) : m/z 286 (13) [M+NH₄]⁺, 269 (100) [M+H]⁺, 268 (22) [M+NH₄-H₂O]⁺, 155 (85) [M-C₅H₅O₃+H]⁺, 114 (6) [M-C₈H₁₁O₃+H]⁺.
- Compound 1b. UV $\lambda_{\max}^{\text{MeOH}}$ nm : 289 . Chromatographic mobilities were recorded in two systems : system 5 (Si gel F-254, CHCl₃:MeOH = 9:1) R_f 0.45, system 6 [LichroCART 250-4 Lichrospher 100 DIOL 5 μm , *n*-Hexane:MeOH:*i*-PrOH = 18:5:2, 1 ml.min⁻¹] R_t 8.5 min. ¹H NMR (500 MHz in MeOH) : δ 0.98 and 1.08 (H-8/9), 2.21 (H-6), 4.64 (H-2). ¹³C NMR (125 MHz in MeOH) : 12.9 (C-6), 16.5 and 17.8 (C-8/9), 42.8 (C-7), 85.2 (C-2), 136.7 (C-4), 174.9 (C-5), 198.4 (C-3). DCI⁺MS (NH₃ + isobutane) (rel. int.) : m/z 286 (13) [M+NH₄]⁺, 269 (100) [M+H]⁺, 268 (22) [M+NH₄-H₂O]⁺, 155 (85) [M-C₅H₅O₃+H]⁺, 114 (6) [M-C₈H₁₁O₃+H]⁺.
- Rodin, J. O., Himel, C. M., Silverstein, Leeper, R. W. and Gortner, W. A. *J. Food Sci.*, **1965**, *30*, 280-285.
- Willhalm, B., Stoll, M. and Thomas, A. F. *Chemistry and Industry* (London), **1965**, *38*, 1629-30.
- Mayerl, F., Näf, R. and Thomas, A. F. *Phytochemistry*, **1989**, *28*, 631-633.